Current Understanding of Pathogenesis and **Treatment of TMJ Osteoarthritis**

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Abstract

Osteoarthritis is a common disease that can cause severe pain and dysfunction in any joint, including the temporomandibular joint (TMJ). TMJ osteoarthritis (TMJOA) is an important subtype in the classification of temporomandibular disorders. TMJOA pathology is characterized by progressive cartilage degradation, subchondral bone remodeling, and chronic inflammation in the synovial tissue. However, the exact pathogenesis and process of TMJOA remain to be understood. An increasing number of studies have recently focused on inflammation and remodeling of subchondral bone during the early stage of TMJOA, which may elucidate the possible mechanism of initiation and progression of TMJOA. The treatment strategy for TMJOA aims at relieving pain, preventing the progression of cartilage and subchondral bone destruction, and restoring joint function. Conservative therapy with nonsteroidal anti-inflammatory drugs, splint, and physical therapy, such as low-energy laser and arthrocentesis, are the most common treatments for TMJOA. These therapies are effective in most cases in relieving the signs and symptoms, but their long-term therapeutic effect on the pathologic articular structure is unsatisfactory. A treatment that can reverse the damage of TMJOA remains unavailable to date. Treatments that prevent the progression of cartilage degradation and subchondral bone damage should be explored, and regeneration for the TMJ may provide the ideal long-term solution. This review summarizes the current understanding of mechanisms underlying the pathogenesis and treatment of TMJOA.

Keywords: temporomandibular joint, temporomandibular disorders, mandibular condyle, bone remodeling, cartilage, inflammation

Introduction

The temporomandibular joint (TMJ) is a synovial joint that performs the most complicated movement in the human body. Osteoarthritis (OA) is a degenerative disease that is characterized by progressive cartilage degradation, subchondral bone remodeling, synovitis, and chronic pain (Zarb and Carlsson 1999). However, the etiology of the majority of TMJ osteoarthritis (TMJOA) is complex and multifactorial or unknown. TMJOA is also an important subtype of temporomandibular disorders (TMDs) (Zarb and Carlsson 1999). It is secondary to disc displacement, trauma, functional overload, and developmental abnormalities, such as secondary TMJOA (Tanaka et al. 2008).

Excessive mechanical loading on normal articular cartilage or normal mechanical loading on impaired articular cartilage is generally speculated to initiate the disruption of cartilage matrix homeostasis, resulting in OA (Tanaka et al. 2008). However, TMJOA may differ from OA in knee or hip, which is closely related to aging, obesity, and overload (Herrero-Beaumont et al. 2009). Overload of the TMJ, including severe malocclusion, skeletal jaw asymmetry, and muscle overuse, has been considered one of the main causes for TMJOA (Tanaka et al. 2008; Matsumoto et al. 2010; Krisjane et al. 2012), but the majority of TMJOA is difficult to attribute to overload. Therefore, the causes of impaired condylar cartilage in the TMJ remain unclear. Increasing attention has focused on

the inflammation and remodeling of subchondral bone, but the pathogenesis of TMJOA remains controversial and unclear.

Patients with TMJOA usually have pain and dysfunction of the TMJ with reduced quality of life. The clinical diagnosis of TMJOA is mainly based on the radiographic features of the condyle and articular eminence, including erosive resorption, sclerosis, attrition, osteophyte formation, and cyst-like change (Zhao et al. 2011; Kalladka et al. 2014). Recently, cone-beam

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Туре	Species	Methods		
Surgical	Rabbit	Partial perforations of the discs	Liu et al. 2011; Zhang et	

Table. Animal Models of Temporomandibular Joint Osteoarthritis Used in the Recent Studies.

Туре	Species	Methods	Applied by
Surgical	Rabbit	Partial perforations of the discs	Liu et al. 2011; Zhang et al. 2011; Xiao et al. 2012; Chen K et al. 2013; Ying et al. 2013
	Rabbit	Defects on the condylar fibrocartilage	Kütük et al. 2014
	Mice	CoII-ILIβXAT	Huang et al. 2013
	Mice	bgn(-/0)fmod(-/-)	Embree et al. 2011
	Mice	Osteoblast-specific mutant TGF- β 1 transgene	Jiao et al. 2014
	Mice	Col9a1(-/-)	Polur et al. 2010
	Mice	Col2αI	Ricks et al. 2013
	Mice	DDR-I(-/-)	Schminke et al. 2014
	Mice	$I\alpha(OH)$ ase(-/-)	Shen et al. 2013
Mechanical	Rat	Experimentally created disordered occlusion	Jiao et al. 2011; Kuang et al. 2013; Zhang J et al. 2013; Zhang M et al. 2013
	Mice	Anterior crossbite prosthesis	Liu et al. 2014
	Rat	Elastic compressive force	Li et al. 2013
Biochemical	Rabbit	Injection of sodium mono iodoacetate (MIA)	Duygu et al. 2011
	Rat	Injection of MIA	Wang et al. 2012; Wang et al. 2013
	Rat	Injection of collagenase	Li et al. 2014
Senescent	Mice	Senescence-accelerated (SAMP8) mice	Ishizuka et al. 2014

computed tomography (CBCT) has provided more detailed change of TMJ bone than conventional radiographic methods (tomography, Schuller's projection, spiral computed tomography, etc.), demonstrating a special advantage in TMJOA diagnosis (dos Anjos Pontual et al. 2012). Treatment of TMJOA is directed at relieving pain, decelerating the progress of the disease, and restoring TMJ function. The pain of patients with TMJOA can be mostly managed effectively with nonsteroidal anti-inflammatory drugs (NSAIDs) or arthrocentesis (Machon et al. 2011). Given the limited understanding of its pathogenesis and the low healing potential of avascular cartilage, no effective therapy is available for restoring the structures of TMJ with progressive OA (de Souza et al. 2012).

Animal models of TMJOA are a critical approach to investigate the pathogenesis of TMJOA and to evaluate the potential therapeutic interventions because obtaining clinical samples from patients with TMJOA is difficult. Several attempts have been made at creating animal models of TMJOA, including surgical, mechanical, chemical, and genetic models. The Table summarizes the animal models used to evaluate the pathogenesis or treatment of TMJOA in recent studies. Although increasing evidence suggests that functional occlusion overload with subsequent microtrauma results in OA-like changes in the TMJ of rodents, anatomical and physiological differences exist between animal model and clinical patients. Most important, the exaggerated occlusion interference in the animal models cannot mimic the condition in the patients with TMJOA. Surgical, transgenic. or chemical methods are also used to induce TMJOA in animals but are still insufficient in mimicking complex clinical conditions. Nevertheless, the animal models of TMJOA will be still irreplaceable at least in the study of the pathology and progression of TMJOA rather than the etiology of OA.

Etiology, diagnosis, and general principles for the therapy of degenerative disease of TMJ, including TMJOA, have been well summarized in previous reviews (Tanaka et al. 2008; de Souza

et al. 2012; Kalladka et al. 2014). This review briefly summarizes the well-known information about TMJOA and emphasizes the current understanding of mechanisms underlying the pathogenesis and treatment of TMJOA published from 2010 to 2014.

Part I. Pathogenesis of TMJOA

Inflammation

TMJOA is classified as a "low-inflammatory arthritic condition," as opposed to rheumatoid arthritis, which is classified as a high-inflammatory condition (de Souza et al. 2012). However, considerable attention has been on the importance of inflammation in the progression of TMJOA. Interleukin (IL)–12 and several other inflammatory cytokines, including IL-1\beta, IL-6, and tumor necrosis factor (TNF)-α, are increased in the synovial fluid of patients with TMJOA (Vernal et al. 2008; Cevidanes et al. 2014). Monocyte chemoattractant protein (MCP)-1 is also elevated in the inflamed synovial tissues and fluids of patients with OA and is highly upregulated in IL-1β– stimulated synoviocytes of the TMJ (Ogura et al. 2010). MCP-1 is speculated to play an important role in recruiting mononuclear cells to inflamed synovial tissues. Expression of IL-1 β and TNF- α is reported to be increased in the experimental chronic inflammation of rodent TMJs, implying that they can be one of the causes for the degenerative changes of TMJ; moreover, the biomechanical property of the disc is decreased in this model (Wang et al. 2014), implying that chronic inflammation in TMJ deteriorates the adaptive capability of the TMJ.

The concentrations of carboxy-terminal telopeptides I and II (CTX-I and CTX-II), serum cartilage oligomeric matrix protein, and prostaglandin E₂ (PGE₂) are higher in the synovial fluid of patients with TMJOA than in the knee joints of patients with OA (Vos et al. 2014). The levels of these markers are not significantly increased in the synovial fluid of patients with TMJOA compared with healthy controls in a previous report (Vos et al. 2013). Whether markers for cartilage degradation are increased in the synovial fluid of patients with TMJOA still needs further investigation. No potential prognostic indicator or diagnostic marker for TMJOA has been identified to date.

Excessive Mechanical Stress

The articular surface is covered by fibrocartilage and the underlying subchondral bone and is stress sensitive and subject to extensive remodeling. Excessive mechanical stress is characterized as a key factor inducing cartilage degradation in the TMJ (Tanaka et al. 2008). A retrospective study shows that unilateral TMJOA is associated with mandibular asymmetry and increasing internal electromyographic activity of the masseter on the OA side (Matsumoto et al. 2010). However, the cause-and-effect relationship among TMJOA, masticatory muscle overuse, and dentofacial morphology remains to be elucidated.

Recent studies have focused on the molecular pathway of articular cartilage degradation and mechanical sensing. The effect of mechanical stress on the mandibular condylar chondrocytes has been evaluated in vitro and in vivo. Excessive mechanical stress induces activation of the plasminogen activator (PA) system, which may lead to proteolysis of extracellular matrix components (Chen W et al. 2013). Long-term experimental disordered occlusion also results in subchondral bone loss and increased osteoclast activation, and the new bone subsequently formed exhibits lower bone mineral density and poor mechanical properties (Zhang J et al. 2013). Other research reported that endoplasmic reticulum stress-induced cell death has an important role in the mandibular cartilage thinning induced by mechanical stress and may be a novel mechanism of chondrocyte apoptosis induced by mechanical force (Li et al. 2013). Several studies explored the molecular mechanisms and signal pathways of mechanical stress-induced TMJOA. Malocclusion-accelerated early abnormal condylar organization in the senescence-accelerated mice is accompanied by a downregulation of Indian hedgehog signaling (Ishizuka et al. 2014). This finding suggests that a possible therapeutic effect of early restoration of hedgehog signaling or that inhibition of endoplasmic reticulum stress can minimize or even prevent the excessive mechanical stress-induced degeneration of TMJs.

Excessive joint loading and abnormal dental occlusion have been used to establish TMJOA models (Jiao et al. 2011). Considerable evidence exists that aberrant biomechanical stimulation possesses an important function in the initiation and progression of TMJOA. However, mechanical stress is also necessary for the development of the mandibular condyle. Female mice (aged 3 wk) with trimmed incisors fed a soft diet for 4 wk were reported to have thinner cartilage and reduced subchondral bone volume in the TMJ (Liu et al. 2014).

Abnormal Remodeling of Subchondral Bone

The clinical diagnosis of TMJOA is mainly based on the radiographic features of the subchondral bone (Kalladka et al. 2014), indicating the important role of subchondral bone in TMJOA. The contribution of the cartilage to the pathology of TMJOA has been studied in depth. An increasing number of studies have recently focused on the effect of subchondral bone on TMJOA pathogenesis (Embree et al. 2011; Jiao et al. 2011; Wang et al. 2012). These studies suggest that increased turnover of subchondral bone plays a role in the initiation or progression of TMJOA.

In the TMJOA model induced by orthodontic disturbed dental occlusion, subchondral bone loss and decreased bone mineral density were observed following degradation of the cartilage. The chondrocytes within the degraded cartilage may regulate osteoclastogenesis by increasing the ratio of the receptor activator of nuclear factor (NF)–κB ligand (RANKL) and osteoprotegerin (OPG) and ultimately result in subchondral bone loss in TMJOA (Jiao et al. 2011). The active interaction of stromal cell–derived factor 1, which binds to its receptor on chondrocytes (CXC chemokine receptor 4) and induces local increases in matrix metalloproteinase (MMP)–9 and IL-6, contributes to the remodeling of subchondral bone in a malocclusion-induced TMJOA model (Kuang et al. 2013).

The upregulation of genes involved in osteoclast activity and an increased RANKL/OPG ratio in subchondral bone likely contribute to the increased subchondral bone turnover of biglycan/fibromodulin-deficient mice during the early stage of TMJOA (Embree et al. 2011). Osteoblast-specific transforming growth factor (TGF)–β1 transgenic mice (aged 4 mo) with high levels of active TGF-β1 in the bone marrow were used to evaluate the effect of overexpressed TGF-β1 on TMJOA (Jiao et al. 2014). In this model, excessive apoptosis of the mandibular condylar chondrocytes; upregulation of MMP-9, MMP-13, and vascular endothelial growth factor (VEGF) in the chondrocytes; and fluctuant bone density of the condyle were observed, implying that TGF-β1 has an initiating role in decreasing bone mineral density and increasing subchondral bone turnover in TMJOA. Remodeling of mandibular condylar subchondral bone is frequently observed in the early stages of TMJOA, but the etiological role of subchondral bone turnover in TMJOA still needs to be further determined.

Chondrocyte Apoptosis

Chondrocyte death caused either by apoptosis or necrosis is assumed to be a central feature in the degeneration of osteoarthritic cartilage clinically or experimentally. In a rat model of iodoacetate-induced TMJOA, the apoptosis of chondrocytes is the prominent characteristic of the early phase of cartilage degradation, and the cytokines released by the apoptotic cartilage chondrocytes may contribute to the destruction of subchondral bone (Wang et al. 2012) (progression of iodoacetate-induced TMJOA in rats is shown in Fig. 1). In malocclusion-induced TMJOA in rat, enhanced chondrocyte autophagy accompanied by a reduction in mitogen-activated protein kinase kinase kinase kinase kinase 3 and mammalian target of rapamycin activity is also observed in the early phase of cartilage degeneration of TMJ (Zhang M et al. 2013). In a senescence-accelerated mouse

model, malocclusion enhances chondrocyte apoptosis in the early stage of cartilage degradation of TMJ (Ishizuka et al. 2014). In addition, 1,25-hydroxyvitamin D (1,25(OH)₂ D)-deficient mice display erosive cartilage degradation in the TMJ by inducing DNA damage, cellular senescence, and the production of senescence-associated inflammatory cytokines, indicating that 1,25(OH), D deficiency may play a role in the pathogenesis of TMJOA (Shen et al. 2013). Oxidative stress induced by H₂O₂ can elevate intracellular reactive oxygen species (ROS) and cause chondrocyte apoptosis and functional impairment in cultured TMJ chondrocytes (Ueno et al. 2011). Although antioxidant amino acid derivative N-acetyl cysteine (NAC) reduces intracellular ROS levels, prevents chondrocyte apoptosis, and increases the expression of aggrecan and type II collagen and production of proteoglycan in vitro (Ueno et al. 2011), the effect of NAC on osteoarthritic chondrocytes remains to be explored in vivo. Taken together, all the current evidence suggests that chondrocyte apoptosis plays an important role in the early phase of TMJOA.

Catabolic Enzymes

The upregulation of catabolic enzymes in the cartilage matrix, such as MMP and a disintegrin and metalloproteinase with thrombos-

pondin motifs (ADAMTS), is involved in the pathology of TMJOA. The expression of ADAMTS-5 is also upregulated in the condylar cartilage in the early stage of TMJOA (Li et al. 2014). Several studies have explored the molecular mechanisms of cartilage degradation because catabolic enzymes catabolize the extracellular matrix.

An in vitro study indicates that the molecular mechanism underlying the IL-1β-induced catabolism of mandibular condylar chondrocytes results from the upregulation of Wnt-5A by activating the NF-κB signaling pathway (Ge et al. 2011). Recently, the expression of high-temperature requirement serine protease A1 (HtrA1) is elevated in the articular cartilage of the TMJ from genetically mutated mouse OA models. This study showed that the upregulation of HtrA1 surrounding the

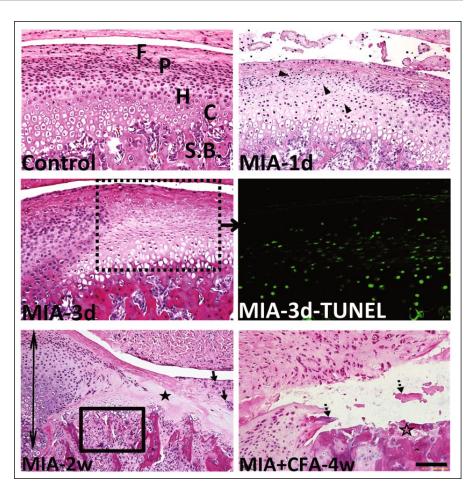


Figure 1. Histopathology of an iodoacetate-induced temporomandibular joint osteoarthritis model in Sprague-Dawley rats. The temporomandibular joint was sectioned in sagittal plane for hematoxylin and eosin staining and terminal deoxynucleotidyl transferase dUTP nick end labeling (TUNEL) assay. Mandibular condyle of each group is shown, including control, MIA-1d (injection of 0.5 mg monosodium iodoacetate [MIA] into the upper compartment of the TMJ for 1 d), MIA-3d (for 3 d), MIA-2w (for 2 wk), and MIA+CFA-4w (combined injection of MIA and complete Freund's adjuvant [CFA, commonly used to induce inflammation] for 4 wk). F, fibrous layer; P, proliferative layer; H, hypertrophy layer; C, calcified layer; S.B., subchondral bone. Dotted black frame is magnified and analyzed by TUNEL, with TUNEL-positive chondrocytes stained green. Star, regional loss of chondrocytes; arrow, chondrocyte cluster formation; double-head arrow, peripheral chondrocyte proliferation; black frame, subchondral bone erosion; dotted arrow, wear-out of cartilage; hollow star, exposure of subchondral bone with dead bone formation. (Bar = 100 μm)

chondrocytes during the early stages of TMJOA initiates the degradation of the chondrocyte pericellular matrix, especially type II collagen (Polur et al. 2010). This may enhance our understanding of the early steps in TMJOA pathogenesis. However, the study used collagen-deficient mice; thus, whether the HtrA1-collagen interaction affects collagen metabolism in nongenetically mutated animal models should be determined.

Estrogen

TMJOA has a female preponderance and occurs mainly after puberty during the reproductive years (Zhao et al. 2011), suggesting a possible function of female hormones in the disease process. Therefore, the effects of estrogen on condylar

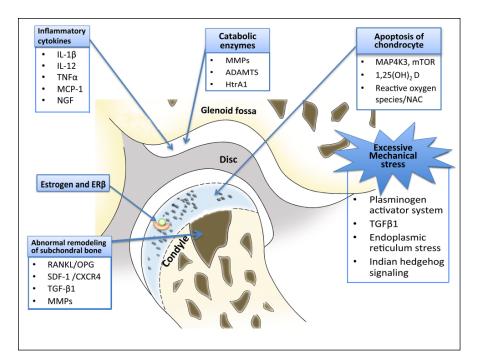


Figure 2. Schematic view of pathogenesis of temporomandibular joint osteoarthritis (TMJOA) reported in recent studies.

cartilage and subchondral bone have been evaluated. Conversion of estrone/17β-estradiol to proinflammatory metabolites can be found in OA synovial cells of the knee joint; this finding implies that proinflammatory metabolites in synoviocytes may be an important mechanism underlying the proinflammatory effects of estradiol in the inflamed TMJ (Schmidt et al. 2009). Estrogen inhibits the mandibular condylar chondrocyte proliferation via an estrogen receptor (ER)-βdependent mechanism in ER-β knockout mice (Chen et al. 2014). Estrogen aggravates the degradation of cartilage and destruction of subchondral bone by upregulating Fas and caspase 3-related proapoptotic genes in an iodoacetate-induced rat model of TMJOA, and these effects of estrogen can be inhibited by an ER antagonist (Wang et al. 2013). These findings suggest that estrogen plays a role in the sexual dimorphism of TMJOA. By contrast, estrogen possesses a protective effect on the TMJ chondrocyte through inhibiting the expression of nitric oxide (Hu et al. 2013). Therefore, the role of estrogen in TMJOA pathogenesis is still inconclusive. Moreover, the effects of other female hormones, including progestin and relaxin, on the progression of cartilage degradation in the TMJ should be evaluated further.

Genetic Factors

Animal models of TMJOA have been established using the transgenic method (Table). Several genes are associated with temporomandibular disorders (Smith et al. 2011). However, a recent genome-wide association study found no single-nucleotide polymorphisms associated with TMJOA diagnosed based mainly

on pain and degenerative bony changes (Yamaguchi et al. 2014). The hypothesis of a genetic susceptibility or predisposition to TMJOA should be evaluated further.

Figure 2 shows the schematic view of TMJOA pathogenesis reported in recent studies.

Part 2. Treatment of TMJOA

TMJOA is a degenerative disease with associated inflammatory changes of the entire joint and can result in severe pain and impaired joint function (Zarb and Carlsson 1999). TMJOA therapy primarily aims to relieve symptoms, stop the disease progress, and restore TMJ function. The traditional treatment for TMJOA mainly includes nonsurgical options, such as physical therapies, occlusal splints, NSAIDs, and arthrocentesis with lubrication or corticosteroid (de Souza et al.

2012). In a review that aimed to evaluate the effect of interventions for the management of TMJOA, de Souza et al. (2012) found an equivalent pain reduction with diclofenac sodium compared with occlusal splints, as well as a similar degree of effectiveness with intra-articular injections that consisted of either sodium hyaluronate or corticosteroid. Some treatment strategies aim to reduce muscle-related overload in the TMJ. A recent retrospective clinical study reports that a stabilization splint is effective in inducing favorable condylar bone remodeling for patients with TMJOA (Ok et al. 2014). A randomized controlled trial (RCT) of 80 patients with TMJOA with singlejoint symptoms suggested that arthrocentesis combined with the use of a splint improves the symptoms (Machon et al. 2011). By contrast, another RCT study on 80 patients with TMD (with only 8% patients with TMJOA) reported that a stabilization splint treatment had no additional benefit in relieving facial pain compared with counseling and masticatory muscle exercises alone in a 1-mo follow-up (Niemelä et al. 2012). Treatment of TMJOA should be directed at eliminating preexisting risk factors. A stabilization splint may be more suitable for the patients with TMJOA with evident muscle overuse or severe bruxism.

Surgery is the last recommendation for the treatment of TMJOA. Surgical intervention, such as joint replacement with autologous bone or an artificial joint, may restore joint function to some extent in severe cases with impaired joint function and intractable pain (Idle et al. 2014). However, joint replacement does not fully restore the destroyed organ, and the long-term prognosis is uncertain, with some cases requiring a second operation. Most of the treatments are effective in terms of decreasing

pain, and some treatments decelerate joint degeneration; however, treatment rarely restores the destroyed joint. RCTs, including participants with a clear diagnosis of TMJOA, should be encouraged to provide high-level evidence for the effectiveness of interventions for the management of TMJOA. The succeeding sections focus on recent studies of TMJOA treatment.

Cytokine-based Therapy

Cartilage is an avascular tissue with low healing potential. Therefore, methods of stimulating the repair of cartilage by intra-articular injection of cytokines or anticytokine have been investigated. Intra-articular injection of IL-1 receptor antagonist or TNF- α inhibitors possesses cartilage-protecting effects in the knee joint (Urech et al. 2010) but has not been attempted to treat TMJOA.

TGF- β_1 promotes the synthesis of extracellular matrix in co-cultures of fibrochondrocytes and chondrocytes (Kalpakci et al. 2011). An in vivo study showed that the intra-articular injection of TGF- β_1 can increase the proteoglycan content in degraded cartilage and prevent damage to subchondral bone in TMJOA induced surgically by bilateral partial perforation of the discs (Ying et al. 2013). The intra-articular injection of NEL-like molecule-1 (Nell-1) into the TMJ with partial discectomy in rabbits upregulates the expression of type II collagen and aggrecan messenger RNAs; this result suggests that the intra-articular injection of Nell-1 is an alternative effective treatment for the cartilage degeneration associated with TMJOA (Xiao et al. 2012).

Platelet-rich plasma (PRP) is blood plasma that contains a large quantity of growth factors released from highly concentrated activated platelets. PRP can improve bone formation in surgically induced severe degenerative changes of TMJ in rabbits but shows no marked effect on repairing cartilage degradation, and the long-term benefits remain unclear (Kütük et al. 2014). Autologous conditioned serum (ACS) as a source of IL-1 receptor antagonist (IL-1Ra) promotes the repair of cartilage and subchondral bone of the knee joint and is an alternative therapy for degenerative joint disease of the knee; however, no reports exist on the use of ACS in the TMJ (Alvarez-Camino et al. 2013). The effect of ACS on TMJOA warrants evaluation because IL-1β also plays an important role in the initiation and progression of TMJOA.

NSAIDs

Upregulation of proinflammatory cytokines is involved in the destruction of the extracellular matrix of the TMOA and also contributes to the chief complaint of TMJ-related pain. The therapeutic effect of NSAIDs is mainly attributed to their inhibition of cyclooxygenase (COX)–2 activity and reduction of cytokine-induced damage to the mandibular condyle. A recent study reported that the selective COX-2 inhibitor celecoxib blocks the upregulation of COX-2, PGE2, aggrecanase, and MMPs and reverses the downregulation of type II collagen and aggrecan in the cultured mandibular condylar chondrocytes

applied with excessive cyclic tensile strain (Su et al. 2014). The protective effects of celecoxib on the homeostasis of mandibular condylar chondrocytes under excessive mechanical stress may explain the mechanism of action of NSAIDs in TMJOA treatment.

Viscosupplementation

Joint lubrication is responsible for maintaining the low-friction environment of the articular surfaces of the TMJ. The coefficient of friction of synovial fluid is higher in patients with TMJOA than in healthy controls (Wei et al. 2010). Hyaluronic acid (HA) is a component of normal synovial fluid and the cartilage matrix of the TMJ. It lubricates the joint, thereby reducing friction and stress on the joint cartilage, and possesses an important function in maintaining TMJ homeostasis. Supplementing the concentration of HA in the joint with exogenous HA via intra-articular injection after arthrocentesis has been proposed as a treatment for joint diseases and is effective in relieving symptoms (Guarda-Nardini et al. 2014). Intra-articular injections of sodium hyaluronate reduce the PA system activity in the synovial fluid of patients with TMJOA, suggesting that the protective effect of HA on TMJOA is associated with regulation of the PA system (Tang et al. 2010). A comparative study demonstrated that intra-articular injection of HA without arthrocentesis is superior to NSAIDs in terms of relieving the symptoms of TMJOA at 1-y follow-up (Triantaffilidou et al. 2013). An animal study also indicates the protective effect of high-molecular-weight HA on iodoacetate-induced TMJOA (Duygu et al. 2011).

However, discrepant findings have been reported. One study reported no protective effect of HA or insulin-like growth factor 1 (IGF-1) alone on cartilage and subchondral bone; IGF-1 enhances proteoglycan synthesis in cartilage and improves the repair of subchondral bone only in combination with HA in TMJOA (Liu et al. 2011). The effect of intra-articular HA injection on the progression of joint damage in OA and the benefit of this treatment remain to be further evaluated.

Regenerative Medicine

Tissue regeneration resulting from cell-based therapy is gaining increasing attention. Regenerative medicine emphasizes the use of stem cells to produce specific tissues. Mesenchymal stem cells (MSCs) are a candidate regenerative therapy for TMJOA because of their ease of collection and ability to differentiate into cartilage and bone. The association of MSCs with human disease suggests novel mechanisms of pathogenesis and possibilities for novel treatments. Bone- and cartilage-like structures were observed after 7 d of in vitro culture of primary human MSCs preconditioned in osteogenic and chondrogenic medium and then seeded in opposite sides of a hyper-hydrated collagen gel (Brady et al. 2011). The injection of stem cells into a defect for regeneration of articular tissues has been proposed (Barry and Murphy 2013). However, such studies mainly focus on OA in the knee; few have evaluated the therapeutic effects of MSCs

in TMJOA. MSCs were reported to survive for at least 4 wk after injection into the upper compartment of the TMJ based on in vivo tracing. The intra-articular injection of MSCs can delay the destruction of cartilage, and the therapeutic effect is enhanced when the MSCs are induced to be chondrogenic in vitro prior to injection (Chen K et al. 2013). This result provides new insights into the role of MSCs in cell-based therapies for TMJOA. However, considerable uncertainty exists regarding the mechanisms and cellular interactions underlying the chondrogenic or osteogenic effects of MSCs in terms of retarding TMJOA progression. Moreover, the chondrogenic potential of MSCs has been reported most frequently in vitro, rarely in vivo. In addition to bone marrow MSCs, oral or dental MSCs should also be tested in TMJOA treatment.

Conclusions

- Proinflammatory cytokines, including IL-1β and TNFα, mediate the imbalance in the metabolism of articular chondrocytes during the progression of TMJOA. Chronic inflammation may deteriorate the adaptive capability of the TMJ.
- 2. Numerous studies have indicated that subchondral bone plays an important role in TMJOA pathology.
- 3. The relationships among inflammation, cartilage erosion, and subchondral bone destruction remain unclear. Mechanical sensing of the articular cartilage and subchondral bone may contribute to the understanding of TMJOA pathogenesis, but associated molecular mechanism and signal pathways are lacking.
- RCTs, including participants with a clear diagnosis of TMJOA, should be encouraged to provide high-level evidence for the effectiveness of interventions for the management of TMJOA.
- Most patients with TMJOA who have pain are treated effectively with NSAIDs or arthrocentesis. Diseasemodifying OA drugs that prevent the progression of cartilage degradation and subchondral bone damage should be further explored.
- 6. Most anticytokine therapies are still in the animal study stage, and clinical trials are necessary.
- 7. Efforts directed toward engineering tissues for repair or replacement of the TMJ will facilitate the development of next-generation treatments and may provide the ideal long-term solution. The regeneration of TMJ cartilage and subchondral bone tissue with suitable mechanical and structural properties represents an attractive new area of research.

Author Contributions

X.D. Wang, contributed to conception, design, and data analysis, drafted and critically revised the manuscript; J.N. Zhang, contributed to data analysis, critically revised the manuscript; Y.H. Gan, Y.H. Zhou, contributed to conception and design, critically revised the manuscript. All authors gave final approval and agree to be accountable for all aspects of the work.

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References

- Alvarez-Camino JC, Vazquez-Delgado E, Gay-Escoda C. 2013. Use of autologous conditioned serum (Orthokine®) for the treatment of the degenerative osteoarthritis of the temporomandibular joint: review of the literature. Med Oral Patol Oral Cir Bucal. 18(3):E433–E438.
- Barry F, Murphy M. 2013. Mesenchymal stem cells in joint disease and repair. Nat Rev Rheumatol. 9(10):584–594.
- Brady MA, Sivananthan S, Mudera V, Liu Q, Wiltfang J, Warnke PH. 2011. The primordium of a biological joint replacement: coupling of two stem cell pathways in biphasic ultrarapid compressed gel niches. J Craniomaxillofac Surg. 39(5):380–386.
- Cevidanes LH, Walker D, Schilling J, Sugai J, Giannobile W, Paniagua B, Benavides E, Zhu H, Marron JS, Jung BT, et al. 2014. 3D osteoarthritic changes in TMJ condylar morphology correlates with specific systemic and local biomarkers of disease. Osteoarthritis Cartilage. 22(10):1657–1667.
- Chen J, Kamiya Y, Polur I, Xu M, Choi T, Kalajzic Z, Drissi H, Wadhwa S. 2014. Estrogen via estrogen receptor beta partially inhibits mandibular condylar cartilage growth. Osteoarthritis Cartilage. 22(11):1861–1868.
- Chen K, Man C, Zhang B, Hu J, Zhu SS. 2013. Effect of in vitro chondrogenic differentiation of autologous mesenchymal stem cells on cartilage and subchondral cancellous bone repair in osteoarthritis of temporomandibular joint. Int J Oral Maxillofac Surg. 42(2):240–248.
- Chen W, Tang Y, Zheng M, Jiang J, Zhu G, Liang X, Li M. 2013. Regulation of plasminogen activator activity and expression by cyclic mechanical stress in rat mandibular condylar chondrocytes. Mol Med Rep 8(4):1155–1162
- de Souza RF, Lovato da Silva CH, Nasser M, Fedorowicz Z, Al-Muharraqi MA. 2012. Interventions for the management of temporomandibular joint osteoarthritis. Cochrane Database Syst Rev. 4:CD007261.
- dos Anjos Pontual ML, Freire JS, Barbosa JM, Frazão MA, dos Anjos Pontual A. 2012. Evaluation of bone changes in the temporomandibular joint using cone beam CT. Dentomaxillofac Radiol. 41(1):24–29.
- Duygu G, Guler N, Cam B, Kürkçü M. 2011. The effects of high molecular weight hyaluronic acid (Hylan G-F 20) on experimentally induced temporomandibular joint osteoartrosis: part II. Int J Oral Maxillofac Surg. 40(12):1406–1413.
- Embree M, Ono M, Kilts T, Walker D, Langguth J, Mao J, Bi Y, Barth JL, Young M. 2011. Role of subchondral bone during early-stage experimental TMJ osteoarthritis. J Dent Res. 90(11):1331–1338.
- Ge XP, Gan YH, Zhang CG, Zhou CY, Ma KT, Meng JH, Ma XC. 2011. Requirement of the NF-kappaB pathway for induction of Wnt-5A by interleukin-1beta in condylar chondrocytes of the temporomandibular joint: functional crosstalk between the Wnt-5A and NF-kappaB signaling pathways. Osteoarthritis Cartilage. 19(1):111–117.
- Guarda-Nardini L, Rossi A, Ramonda R, Punzi L, Ferronato G, Manfredini D. 2014. Effectiveness of treatment with viscosupplementation in temporomandibular joints with or without effusion. Int J Oral Maxillofac Surg. 43(10):1218–1223.
- Herrero-Beaumont G, Roman-Blas JA, Castaneda S, Jimenez SA. 2009. Primary osteoarthritis no longer primary: three subsets with distinct etio-logical, clinical, and therapeutic characteristics. Semin Arthritis Rheum. 39(2):71–80.
- Hu F, Zhu W, Wang L. 2013. MicroRNA-203 up-regulates nitric oxide expression in temporomandibular joint chondrocytes via targeting TRPV4. Arch Oral Biol. 58(2):192–199.
- Huang H, Shank G, Ma L, Tallents RH, Kyrkanides S. 2013. Nerve growth factor induced after temporomandibular joint inflammation decelerates chondrocyte differentiation. Oral Dis. 19(6):604–610.
- Idle MR, Lowe D, Rogers SN, Sidebottom AJ, Speculand B, Worrall SF. 2014. UK temporomandibular joint replacement database: report on baseline data. Br J Oral Maxillofac Surg. 52(3):203–207.
- Ishizuka Y, Shibukawa Y, Nagayama M, Decker R, Kinumatsu T, Saito A, Pacifici M, Koyama E. 2014. TMJ degeneration in SAMP8 mice is accompanied by deranged Ihh signaling. J Dent Res. 93(3):281–287.

- Jiao K, Niu LN, Wang MQ, Dai J, Yu SB, Liu XD, Wang J. 2011. Subchondral bone loss following orthodontically induced cartilage degradation in the mandibular condyles of rats. Bone. 48(2):362–371.
- Jiao K, Zhang M, Niu L, Yu S, Zhen G, Xian L, Yu B, Yang K, Liu P, Cao X, et al. 2014. Overexpressed TGF-beta in subchondral bone leads to mandibular condyle degradation. J Dent Res. 93(2):140–147.
- Kalladka M, Quek S, Heir G, Eliav E, Mupparapu M, Viswanath A. 2014. Temporomandibular joint osteoarthritis: diagnosis and long-term conservative management: a topic review. J Indian Prosthodont Soc. 14(1):6–15.
- Kalpakci KN, Kim EJ, Athanasiou KA. 2011. Assessment of growth factor treatment on fibrochondrocyte and chondrocyte co-cultures for TMJ fibrocartilage engineering. Acta Biomater. 7(4):1710–1718.
- Krisjane Z, Urtane I, Krumina G, Neimane L, Ragovska I. 2012. The prevalence of TMJ osteoarthritis in asymptomatic patients with dentofacial deformities: a cone-beam CT study. Int J Oral Maxillofac Surg. 41(6):690–695.
- Kuang B, Dai J, Wang QY, Song R, Jiao K, Zhang J, Tian XG, Duan YZ, Wang MQ. 2013. Combined degenerative and regenerative remodeling responses of the mandibular condyle to experimentally induced disordered occlusion. Am J Orthod Dentofacial Orthop. 143(1):69–76.
- Am J Orthod Dentofacial Orthop. 143(1):69–76. Kütük N, Baş B, Soylu E, Gönen ZB, Yilmaz C, Balcioğlu E, Özdamar S, Alkan A. 2014. Effect of platelet-rich plasma on fibrocartilage, cartilage, and bone repair in temporomandibular joint. J Oral Maxillofac Surg. 72(2):277–284.
- Li H, Zhang XY, Wu TJ, Cheng W, Liu X, Jiang TT, Wen J, Li J, Ma QL, Hua ZC. 2013. Endoplasmic reticulum stress regulates rat mandibular cartilage thinning under compressive mechanical stress. J Biol Chem. 288(25):18172–18183.
- Li W, Wu M, Jiang S, Ding W, Luo Q, Shi J. 2014. Expression of ADAMTs-5 and TIMP-3 in the condylar cartilage of rats induced by experimentally created osteoarthritis. Arch Oral Biol. 59(5):524–529.
- Liu XW, Hu J, Man C, Zhang B, Ma YQ, Zhu SS. 2011. Insulin-like growth factor-1 suspended in hyaluronan improves cartilage and subchondral cancellous bone repair in osteoarthritis of temporomandibular joint. Int J Oral Maxillofac Surg. 40(2):184–190.
- Liu YD, Liao LF, Zhang HY, Lu L, Jiao K, Zhang M, Zhang J, He JJ, Wu YP, Chen D, et al. 2014. Reducing dietary loading decreases mouse temporomandibular joint degradation induced by anterior crossbite prosthesis. Osteoarthritis Cartilage. 22(2):302–312.
- Machon V, Hirjak D, Lukas J. 2011. Therapy of the osteoarthritis of the temporomandibular joint. J Craniomaxillofac Surg. 39(2):127–130.
- Matsumoto R, Ioi H, Goto TK, Hara A, Nakata S, Nakasima A, Counts AL. 2010. Relationship between the unilateral TMJ osteoarthritis/osteoarthrosis, mandibular asymmetry and the EMG activity of the masticatory muscles: a retrospective study. J Oral Rehabil. 37(2):85–92.
- Niemelä K, Korpela M, Raustia A, Ylöstalo P, Sipilä K. 2012. Efficacy of stabilisation splint treatment on temporomandibular disorders. J Oral Rehabil. 39(11):799–804.
- Ogura N, Satoh K, Akutsu M, Tobe M, Kuyama K, Kuboyama N, Sakamaki H, Kujiraoka H, Kondoh T. 2010. MCP-1 production in temporomandibular joint inflammation. J Dent Res. 89(10):1117–1122.
- Ok SM, Lee J, Kim YI, Lee JY, Kim KB, Jeong SH. 2014. Anterior condylar remodeling observed in stabilization splint therapy for temporoman-dibular joint osteoarthritis. Oral Surg Oral Med Oral Pathol Oral Radiol. 118(3):363–370.
- Polur I, Lee PL, Servais JM, Xu L, Li Y. 2010. Role of HTRA1, a serine protease, in the progression of articular cartilage degeneration. Histol Histopathol. 25(5):599–608.
- Ricks ML, Farrell JT, Falk DJ, Holt DW, Rees M, Carr J, Williams T, Nichols BA, Bridgewater LC, Reynolds PR, et al. 2013. Osteoarthritis in temporomandibular joint of Col2a1 mutant mice. Arch Oral Biol. 58(9):1092–1099.
- Schmidt M, Hartung R, Capellino S, Cutolo M, Pfeifer-Leeg A, Straub RH. 2009. Estrone/17beta-estradiol conversion to, and tumor necrosis factor inhibition by, estrogen metabolites in synovial cells of patients with rheumatoid arthritis and patients with osteoarthritis. Arthritis Rheum. 60(10):2913–2922.
- Schminke B, Muhammad H, Bode C, Sadowski B, Gerter R, Gersdorff N, Bürgers R, Monsonego-Ornan E, Rosen V, Miosge N. 2014. A discoidin domain receptor 1 knock-out mouse as a novel model for osteoarthritis of the temporomandibular joint. Cell Mol Life Sci. 71(6):1081–1096.
- Shen M, Luo Y, Niu Y, Chen L, Yuan X, Goltzman D, Chen N, Miao D. 2013. 1,25(OH)(2)D deficiency induces temporomandibular joint osteoarthritis via secretion of senescence-associated inflammatory cytokines. Bone. 55(2):400–409.

- Smith SB, Maixner DW, Greenspan JD, Dubner R, Fillingim RB, Ohrbach R, Knott C, Slade GD, Bair E, Gibson DG, et al. 2011. Potential genetic risk factors for chronic TMD: genetic associations from the OPPERA case control study. J Pain. 12(11 Suppl):T92–101.
- Su SC, Tanimoto K, Tanne Y, Kunimatsu R, Hirose N, Mitsuyoshi T, Okamoto Y, Tanne K. 2014. Celecoxib exerts protective effects on extracellular matrix metabolism of mandibular condylar chondrocytes under excessive mechanical stress. Osteoarthritis Cartilage. 2(6):845–851.
- Tanaka E, Detamore MS, Mercuri LG. 2008. Degenerative disorders of the temporomandibular joint: etiology, diagnosis, and treatment. J Dent Res. 87(4):296–307.
- Tang YL, Zhu GQ, Hu L, Zheng M, Zhang JY, Shi ZD, Liang XH. 2010. Effects of intra-articular administration of sodium hyaluronate on plasminogen activator system in temporomandibular joints with osteoarthritis. Oral Surg Oral Med Oral Pathol Oral Radiol Endod. 109(4):541–547.
- Triantaffilidou K, Venetis G, Bika O. 2013. Efficacy of hyaluronic acid injections in patients with osteoarthritis of the temporomandibular joint: a comparative study. J Craniofac Surg. 24(6):2006–2009.
- Ueno T, Yamada M, Sugita Y, Ogawa T. 2011. N-acetyl cysteine protects TMJ chondrocytes from oxidative stress. J Dent Res. 90(3):353–359.
- Urech DM, Feige U, Ewert S, Schlosser V, Ottiger M, Polzer K, Schett G, Lichtlen P. 2010. Anti-inflammatory and cartilage-protecting effects of an intra-articularly injected anti-TNFα single-chain Fv antibody (ESBA105) designed for local therapeutic use. Ann Rheum Dis. 69(2):443–449.
- Vernal R, Velásquez E, Gamonal J, Garcia-Sanz JA, Silva A, Sanz M. 2008. Expression of proinflammatory cytokines in osteoarthritis of the temporomandibular joint. Arch Oral Biol. 53(10):910–915.
- Vos LM, Kuijer R, Huddleston Slater JJ, Bulstra SK, Stegenga B. 2014. Inflammation is more distinct in temporomandibular joint osteoarthritis compared to the knee joint. J Oral Maxillofac Surg. 72(1):35–40.
- Vos LM, Kuijer R, Huddleston Slater JJ, Stegenga B. 2013. Alteration of cartilage degeneration and inflammation markers in temporomandibular joint osteoarthritis occurs proportionally. J Oral Maxillofac Surg. 71(10):1659–1664.
- Wang XD, Cui SJ, Liu Y, Luo Q, Du RJ, Kou XX, Zhang JN, Zhou YH, Gan YH. 2014. Deterioration of mechanical properties of discs in chronically inflamed TMJ. J Dent Res. 93(11):1170–1176.
- Wang XD, Kou XX, He DQ, Zeng MM, Meng Z, Bi RY, Liu Y, Zhang JN, Gan YH, Zhou YH. 2012. Progression of cartilage degradation, bone resorption and pain in rat temporomandibular joint osteoarthritis induced by injection of iodoacetate. PLoS One. 7(9):e45036.
- Wang XD, Kou XX, Meng Z, Bi RY, Liu Y, Zhang JN, Zhou YH, Gan YH. 2013. Estrogen aggravates iodoacetate-induced temporomandibular joint osteoarthritis. J Dent Res. 92(10):918–924.
- Wei L, Xiong H, Li B, Cheng Y, Long X. 2010. Boundary-lubricating ability and lubricin in synovial fluid of patients with temporomandibular joint disorders. J Oral Maxillofac Surg. 68(10):2478–2483.
- Xiao D, Hu J, Chen K, Man C, Zhu S. 2012. Protection of articular cartilage by intra-articular injection of NEL-like molecule 1 in temporomandibular joint osteoarthritis. J Craniofac Surg. 23(1):e55–e58.
- Yamaguchi T, Nakaoka H, Yamamoto K, Fujikawa T, Kim YI, Yano K, Haga S, Katayama K, Shibusawa T, Park SB, et al. (2014). Genome-wide association study of degenerative bony changes of the temporomandibular joint. Oral Dis. 20(4):409–415.
- Ying B, Chen K, Hu J, Man C, Feng G, Zhang B, Zhu S. 2013. Effect of different doses of transforming growth factor-beta(1) on cartilage and subchondral bone in osteoarthritic temporomandibular joints. Br J Oral Maxillofac Surg. 51(3):241–246.
- Zarb GA, Carlsson GE. 1999. Temporomandibular disorders: osteoarthritis. J Orofac Pain. 13(4):295–306.
- Zhang B, Hu J, Man C, Zhu S. 2011. Effect of intra-articular administration of interleukin 1 receptor antagonist on cartilage repair in temporomandibular joint. J Craniofac Surg. 22(2):711–714.
- Zhang J, Jiao K, Zhang M, Zhou T, Liu XD, Yu SB, Lu L, Jing L, Yang T, Zhang Y, et al. 2013. Occlusal effects on longitudinal bone alterations of the temporomandibular joint. J Dent Res. 92(3):253–259.
- Zhang M, Zhang J, Lu L, Qiu ZY, Zhang X, Yu SB, Wu YP, Wang MQ. 2013. Enhancement of chondrocyte autophagy is an early response in the degenerative cartilage of the temporomandibular joint to biomechanical dental stimulation. Apoptosis. 18(4):423–434.
- Zhao YP, Zhang ZY, Wu YT, Zhang WL, Ma XC. 2011. Investigation of the clinical and radiographic features of osteoarthrosis of the temporomandibular joints in adolescents and young adults. Oral Surg Oral Med Oral Pathol Oral Radiol Endod. 111(2):e27–e34.